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# Enhancing Drought Stress Tolerance in Prilep Tobacco Plants with Melatonin Treatment

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Author's contribution

The sole author designed, analysed, interpreted and prepared the manuscript.

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Original Research Article

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### ABSTRACT

In this work, we studied the effect of treatment with melatonin at concentrations (50, 100 and 150 micromolar) on some biochemical and production characteristics of Prilep tobacco plants under conditions of applied drought stress (15%, 30% and 45%).

Chlorophyll content in leaves decreased under conditions of drought stress, and  $H_2O_2$ , proline, MDA and Total Soluble Sugars increased steadily with increasing applied stress, while Chlorophyll, total soluble sugars content in leaves increased when sprayed with melatonin, especially at low concentrations (50 micromolar).

The treatment with the reduced concentration of melatonin and the applied stress outperformed all treatments and the control for all indicators studied. Therefore, it is recommended to use melatonin, especially at a concentration of 50 micromolar, on tobacco plants because of its role in improving chemical traits under conditions of drought stress.

Keywords: Tobacco seeds; Prilep; melatonin; drought stress.

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### **1. INTRODUCTION**

"Abiotic stress greatly limits the growth and yield of plants. With the increase in global temperatures, the effect of drought stress is particularly prominent, and has become the main environmental factor limiting the growth and development of plants worldwide" [1].

"Drought stress increases the production of reactive oxygen species (ROS) such as  $H_2O_2$ . Membrane lipid peroxidation is triggered when the balance between ROS accumulation and the free radical scavenging systems is disrupted, causing damage to the membrane system and leading to an imbalance in plant metabolism" [2].

Drought stress has been shown to have a harmful effect on plant performance [3] including plant growth [4], leaf water content [5] and redox characteristics [6].

"Drought is among the most critical environmental concerns in agriculture today, especially in dry and semi-dry climates including the Mediterranean area" [7]. "Plant biologists have focused their efforts on studying the deleterious effects of drought on plants as well as techniques for mitigating those effects" [8].

"It has been reported that under water deficit stress, compared with no treatment, the application of exogenous hormones can significantly alleviate growth and developmental impairments and improve the activity of antioxidant enzymes" [9].

Melatonin (N-acetyl-5 methoxytryptamine), also known as the pineal hormone [10], was first detected in plants by Hattori [11].

"One important function of melatonin is as an antioxidant that can alleviate plant damage under adverse conditions. Such antioxidant effects have been reported in several plants (e.g., maize, wheat" [12,13].

"Research has shown that under water stress, melatonin can directly scavenge ROS and also improve the activity of antioxidant enzymes to alleviate the oxidative damage caused by water stress in cells" [14].

Therefore, this research aims to improve the biochemical properties of tobacco plants (Prilep) by foliar spraying with melatonin when exposed to drought stress conditions.

### 2. MATERIALS AND METHODS

The experiment was carried out during the 2024 season. The field experiment was conducted in the village of Jinghil- Lattakia- Syria.

Tobacco seeds were grown on an agricultural medium containing compost. The seedlings were transferred to plastic bags ( $60 \times 40$ ) cm forty days after germination.

**Melatonin treatments:** Melatonin solution was prepared by dissolving melatonin in ethanol (50 mg of melatonin in 1 ml of ethanol), after preparing the solution, it was diluted with deionized water to obtain concentrations (0, 50, 100 and 150) micromolar for use in foliar spraying for four days in the dark [15].

**Drought stress treatments:** To induce drought stress, polyethylene glycol (PEG-6000) was used at concentrations (15, 30 and 45 %) equivalent to an osmotic pressure (-0.7, -1.4 and -2.1) MPa [16]. PEG 6000 causes obvious water stress to plants without any toxic effects [17]. Through irrigation at a rate of two irrigations of 200 ml per plant at each treatment, Irrigation was done to the bottom of the plant stem, with an interval of two weeks between one irrigation and the second, during the critical growth period of the plant, which corresponds to the stage of active vegetative growth, about a month after transplanting.

- D<sub>0</sub>: Plants were irrigated with water only.
- D<sub>1</sub>: The plants were irrigated with a solution at a concentration of 15%, equivalent to an osmotic pressure of -0.7 MPa
- D<sub>2</sub>: The plants were irrigated with a 30% solution equivalent to an osmotic pressure of -1.4 MPa.
- D<sub>3</sub>: The plants were irrigated with a solution at a concentration of 45%, equivalent to an osmotic pressure of -2.1 MPa.

Observations were recorded *viz*, Chlorophyll and carotenoid,  $H_2O_2$ , proline, MDA and sugar content in leaves.

## • Determination of Chlorophyll content in leaves

Chlorophyll content determination Pigments were extracted from the leaves. Th e extraction of leaf pigments was performed with 80% acetone, and the absorbance at 663 and 645 nm was measured with an Amersham spectrophotometer (Amersham Biosciences, Piscataway, NJ, USA). The total chlorophyll quantities were calculated according to the method of Arnon [18]. Total carotenoid content was measured at 470 nm. The pigment concentrations were expressed as  $\mu g g^{-1}$  fresh weight (FW).

#### • Determination of H<sub>2</sub>O<sub>2</sub> content in leaves

The  $H_2O_2$  level was measured colorimetrically as described by Jana and Choudhuri [19].  $H_2O_2$  was extracted by homogenizing 50 mg leaf tissue with 3 mL of phosphate buffer (50 mM, pH 6.5).

## • Determination of proline content in leaves:

The proline content was determined according to Bates et al. [20]. "Frozen leaf tissue (0.5 g) was homogenized with 10 mL of 3% sulfosalicylic acid at 4 °C. Th e extract was filtered with Whatman No. 2 filter paper. In a test tube, 2 mL of fi Itrate, 2 mL of acid-ninhydrin, and 2 mL of glacial acetic acid were mixed and incubated at 100 °C for 1 h. Th e reaction was terminated on ice, and the reaction mixture was then extracted with 4 mL of toluene. The chromophorecontaining toluene was separated from the hydrated phase. The absorbance at 520 nm was spectrophotometrically determined with toluene as the blank. The proline concentration was calculated based on a standard curve and was expressed as µmol proline g<sup>-1</sup> FW". Bates et al. [20].

## • Determination of malondialdehyde content in leaves:

"Extract the MDA from 50 mg plant material using 1 ml 0.25% thiobarbituric acid (TBA) dissolved in 10% trichloroacetic acid (TCA). 2. Collect the extract in a 1.5-ml eppendorf tube, heat the mixture at 85 C for 30 min, and then guickly chill it on ice. Proceed by the same way for the blank sample but without plant material. 3. Centrifuge the mixture at maximum speed for 10 min to pellet the particles. 4. Transfer the supernatant in a 1-ml plastic cuvette for spectrophotometric measurement. 5. Read the absorbance first at 532 nm (the peak of MDA-TBA complex) and second time at 600 nm (nonspecific absorption). As blank, use 1 ml 0.25% TBA in 10% TCA. Calculate A (532-600)" (Heath and Packer, 1968). 6. Estimate the MDA concentration using the Beer-Lambert-Bouguer law, MDA extinction coefficient e532 155 mM1 cm1, calculate the amount of MDA, and normalize the values to the fresh weight of each sample.

## • Determination of total soluble sugars in leaves (%):

Quantification of Sugars Samples for sugar determinations were harvested in the morning, between 3 and 4 h into the photoperiod. Sugars were extracted in 80% ethanol and determined enzymatically as described by Scholes *et al.* [21].

**Statistical Analysis:** Statistical analyses were performed by the analysis of variance (ANOVA) with Tukey. All data are presented as means  $\pm$  standard deviation (SD) of three replicates. Differences at P < 0.05 were considered to be significant.

### 3. RESULTS AND DISCUSSION

Effect of melatonin and drought stress on chlorophyll content in leaves: Data in Fig. 1 indicate that there are significant differences (P<0.05) between the studied treatments in terms of the chlorophyll content in leaves.

Drought stress led to a decrease in chlorophyll content in leaves, While treatment with melatonin increased chlorophyll content in leaves compared to the control.

Treatment with melatonin at a concentration of 50 micromolar under drought conditions also outperformed all other parameters and the control.

"In general, drought stress causes a series of physiological and biochemical reactions such as stomatal closure, decreased chlorophyll content, decreased transpiration, and reduced antioxidant capacity" [22].

"Drought stress inhibits plant growth and yield by reducing its photosynthetic capacity and results in an imbalance in the cellular redox modules, with antioxidant defences failing to counterbalance the increased creation of reactive oxygen species (ROS)" [23]. "This leads a cascade of oxidative damage, impairing development and growth" [24] melatonin inhibit chlorophyll degradation, and increase leaf photosynthetic rates and the accumulation of dry matter [25].

Effect of melatonin and drought stress on H2O2 content in leaves: Data in Fig. 2 indicate that there are significant differences (P<0.05) between the studied treatments in terms of the H<sub>2</sub>O<sub>2</sub> content in leaves.

Drought stress led to an increase in  $H_2O_2$  content in leaves, While treatment with melatonin decreased  $H_2O_2$  content in leaves compared to the control.

Treatment with melatonin under drought conditions at low concentration (50 micromolar) also led to an decrease in the  $H_2O_2$  compared to the remaining treatments and the control.

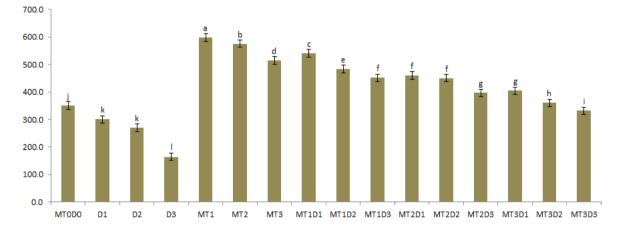
"Stress increases the active oxygen yield and causes damage to cells. The H<sub>2</sub>O<sub>2</sub> is an important signaling molecule for the plant stress

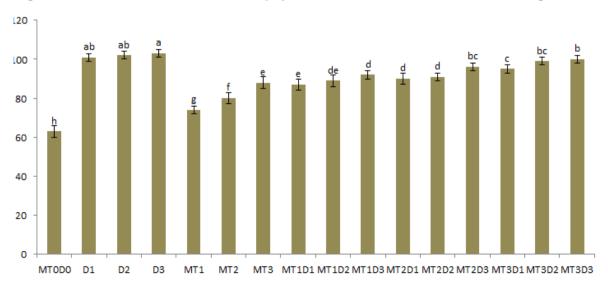
response, and is widely involved in plant physiological and cross-resistance processes" [26].

Therefore, melatonin can directly scavenge excess ROS [27], and it can also promote the accumulation of other antioxidants and antioxidant enzymes to indirectly remove ROS [28].

Effect of melatonin and drought stress on proline content in leaves: Data in Fig. 3 indicate that there are significant differences (P<0.05) between the studied treatments in terms of the proline content in leaves.

drought stress led to an increase in proline content in leaves, While treatment with melatonin decreased proline content in leaves compared to the control.





#### Fig. 1. Effect of melatonin on the chlorophyll content in tobacco leaves under drought stress

Fig. 2. Effect of melatonin on the H<sub>2</sub>O<sub>2</sub> content in tobacco leaves under drought stress

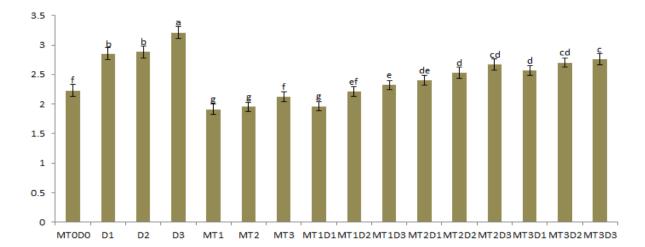


Fig. 3. Effect of melatonin on the proline content in tobacco leaves under drought stress

Treatment with melatonin under drought conditions at low concentration (50 micromolar) also led to an decrease in the proline compared to the remaining treatments and the control.

The results of current study showed that proline concentrations increased in response to drought, which is in agreement with prior research conducted by Cao et al. [29].

"This finding supports the notion that increased accumulation of free amino acids and proline is a common response of plants to water deficit conditions. Proline also protects plant cells from oxidative damage and effectively neutralises ROS in addition to acting as an osmoprotectant" [30]. Effect of melatonin and drought stress on malondialdehyde content in leaves: Data in Fig. 4 indicate that there are significant differences (P<0.05) between the studied treatments in terms of the malondialdehyde content in leaves.

drought stress led an increase in to malondialdehyde content leaves, While in treatment with melatonin decreased malondialdehyde content in leaves compared to the control.

Treatment with melatonin under drought conditions at low concentration also led to an decrease in the malondialdehyde compared to the remaining treatments and the control.

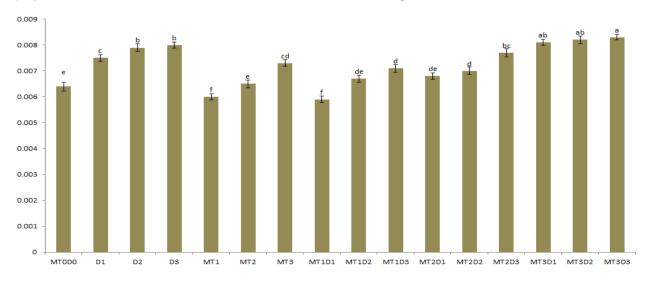
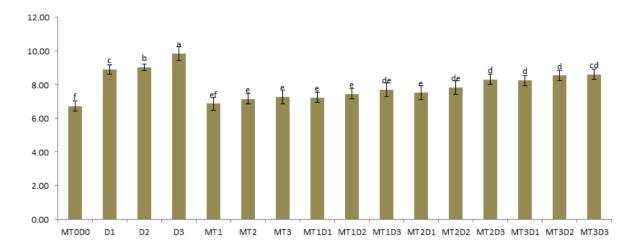


Fig. 4. Effect of melatonin on the malondialdehyde content in tobacco leaves under drought stress



## Fig. 5. Effect of melatonin on the malondialdehyde content in tobacco leaves under drought stress

"MDA and  $H_2O_2$  are well-known oxidative damage metrics and both were elevated under drought in the current study, indicating increased oxidative stress in drought-treated plants" [31].

"MT pre-treatments alleviated oxidative stress in drought-treated plants, as indicated by lower  $H_2O_2$  and MDA levels. Additional elevations in GST were achieved with the pre-treatment of MT under drought, showing that GST plays a positive function in detoxifying  $H_2O_2$ " [32].

"This confirms the role of MT as an inducer of antioxidant enzymatic activity and eliminating ROS. Melatonin is a multifunctional molecule that is known as an all-purpose antioxidant per se" [33].

Effect of melatonin and drought stress on sugar content in leaves: Data in Fig. 5 indicate that there are significant differences (P<0.05) between the studied treatments in terms of the sugar content in leaves.

drought stress led to an increase in sugar content in leaves, While treatment with melatonin decreased sugar content in leaves compared to the control.

Treatment with melatonin under drought conditions at low concentration also led to an decrease in the sugar compared to the remaining treatments and the control.

"Osmolytes, such as proline and soluble sugars, are produced by plants due to drought stress to

maintain high osmotic pressure and water potential within the cells, maintaining the integrity of the cell membranes and enhancing drought resistance in crops" [34].

The accumulation and synthesis of soluble compounds, such as amino acids, particularly proline and soluble sugars, control osmoregulation during drought [35-37].

#### 4. CONCLUSIONS

The study concluded that the application of melatonin can stimulate the antioxidant defense system, lower ROS levels. According to the results, using exogenous melatonin can be a viable and affordable way to lower crop losses and raise potential yields in water-limited environments. In order to increase plant tolerance and drought adaptation, our study also emphasizes the necessity of more research into the molecular mechanisms underpinning melatonin action and its effect on nutrient absorption. In light of global warming, the development of low-cost, environmentally friendly technologies, such as melatonin application, is critical. Our research offers fresh perspectives on the potential uses of melatonin in crop production.

#### DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts.

#### **COMPETING INTERESTS**

Author has declared that no competing interests exist.

### REFERENCES

- Qi J, Song CP, Wang B, Zhou J, Kangasjärvi J, Zhu JK, Gong Z. Reactive oxygen species signaling and stomatal movement in plant responses to drought stress and pathogen attack. J Integr Plant Biol. 2018;60(9):805-26.
- Nahar K, Hasanuzzaman M, Alam MM, Fujita MJ. Roles of exogenous glutathione in antioxidant defense system and methylglyoxal detoxification during salt stress in mung bean. Biol Plant. 2015;59:745-56.
- Khan N, Bano A, Babar MA. Metabolic and physiological changes induced by plant growth regulators and plant growth promoting rhizobacteria and their impact on drought tolerance in *Cicer arietinum* L. PLoS One. 2019;14(3).
- Al-Huqail A, El-Dakak RM, Sanad MN, Badr RH, Ibrahim MM, Soliman D, Khan F. Effects of climate temperature and water stress on plant growth and accumulation of antioxidant compounds in sweet basil (*Ocimum basilicum* L.) leafy vegetable. Scientifica (Cairo). 2020;2020:3808909.
- Ichsan CN, Basyah B, Zakaria S, Efendi E. Differences of water status and relationship with roots growth and yield of rice under water stress. Syst Rev Pharm. 2020;11(8).
- Bernardo L, Carletti P, Badeck FW, Rizza F, Morcia C, Ghizzoni R, Rouphael Y, Colla G, Terzi V, Lucini L. Metabolomic responses triggered by arbuscular mycorrhiza enhance tolerance to water stress in wheat cultivars. Plant Physiol Biochem. 2019;137:203-12.
- Huang Z, Liu X, Sun S, Tang Y, Yuan X, Tang Q. Global assessment of future sectoral water scarcity under adaptive inner-basin water allocation measures. Sci Total Environ. 2021;783:146973.
- Ahmad S, Wang GY, Muhammad I, Chi YX, Zeeshan M, Nasar J, Zhou XB. Interactive effects of melatonin and nitrogen improve drought tolerance of maize seedlings by regulating growth and physiochemical attributes. Antioxidants (Basel). 2022;11(2):359.

- Wu YY, Tian YD, Liu LX, Xing XH, Jiang HQ, Xing H, Zhou Q. Effects of Triadimefon on physiological characteristics and yield of soybean under drought and rewatering at flowering stage. J Nucl Agric Sci. 2013;27:1749 -55.
- Arnao MB, Hernández-Ruiz J. Functions of melatonin in plants: a review. J Pineal Res. 2015;59(2):133-50.
- 11. Dubbels R, Reiter RJ, Klenke E, Goebel A, Schnakenberg E, Ehlers C, Schloot W. Melatonin in edible plants identified by radioimmunoassay and by high performance liquid chromatography-mass spectrometry. J Pineal Res. 1995;18(1):28-31.
- Chen YE, Mao JJ, Sun LQ, Huang B, Ding CB, Gu Y, Yuan M. Exogenous melatonin enhances salt stress tolerance in maize seedlings by improving antioxidant and photosynthetic capacity. Physiol Plant. 2018;164(3):349-63.
- Turk H, Erdal S, Genisel M, Atici O, Demir Y, Yanmis D. The regulatory effect of melatonin on physiological, biochemical and molecular parameters in cold-stressed wheat seedlings. Plant Growth Regul. 2014;74:139-52.
- Zhang N, Zhao B, Zhang HJ, Weeda S, Yang C, Yang ZC, Guo YD. Melatonin promotes water-stress tolerance, lateral root formation, and seed germination in cucumber (*Cucumis sativus* L.). J Pineal Res. 2013;54(1):15-23.
- Liu L, Li D, Ma YL, Wang LJ, Zhao SM, Zhou JX, Shen HT, Wang YF. Alleviation of drought stress and the physiological mechanisms in tobacco seedlings treated with exogenous melatonin. Acta Prataculturae Sinica. 2019;28(8):95.
- Muscolo A, Sidari M, Anastasi U, Santonoceto C, Maggio A. Effect of PEGinduced drought stress on seed germination of four lentil genotypes. J Plant Interact. 2014;9(1):354-63.
- 17. Emmerich WE, Hardegree SP. Polyethylene glycol solution contact effects on seed germination. Agron J. 1990;82(6):1103-7.
- Arnon DI. Copper enzymes in isolated chloroplasts. Polyphenoloxidase in Beta vulgaris. Plant Physiol. 1949;24(1):1.
- 19. Jana S, Choudhuri MA. Glycolate metabolism of three submersed aquatic angiosperms during ageing. Aquat Bot. 1982;12:345-54.

- 20. Bates LS, Waldren RP, Teare ID. Rapid determination of free proline for waterstress studies. Plant Soil. 1973;39:205-7.
- 21. Scholes JD, Lee PJ, Horton P, Lewis DH. Invertase: understanding changes in the photosynthetic and carbohydrate metabolism of barley leaves infected with powdery mildew. New Phytol. 1994; 126(2):213-22.
- 22. Farooq M, Wahid A, Kobayashi NSMA, Fujita DBSMA, Basra SM. Plant drought stress: effects, mechanisms and management. Sustainable Agriculture. 2009:153-88.
- Sun Y, Li Y, Wang M, Wang C, Ling N, Mur LA, Shen Q, Guo S. Redox imbalance contributed differently to membrane damage of cucumber leaves under water stress and Fusarium infection. Plant Sci. 2018;274:171-80.
- Kerchev PI, Van Breusegem F. Improving oxidative stress resilience in plants. Plant J. 2022;109(2):359-72.
- Arnao MB, Hernández-Ruiz J. Protective effect of melatonin against chlorophyll degradation during the senescence of barley leaves. J Pineal Res. 2009; 46(1):58-63.
- Apel K, Hirt H. Reactive oxygen species: metabolism, oxidative stress, and signal transduction. Annu Rev Plant Biol. 2004;55(1):373-99.
- 27. Reiter RJ, Tan DX, Burkhardt S, Manchester LC. Melatonin in plants. Nutr Rev. 2001;59(9):286-90.
- Zhang N, Zhang H, Yang R, Huang Y, Guo Y. Advances in melatonin and its functions in plants. Agric Sci Technol. 2012;13(9):1833.
- 29. Cao L, Qin B, Gong Z, Zhang Y. Melatonin improves nitrogen metabolism during grain filling under drought stress. Physiol Mol Biol Plants. 2022;28(7):1477-88.

- Ould said C, Boulahia K, Eid MA, Rady MM, Djebbar R, Abrous-Belbachir O. Exogenously used proline offers potent antioxidative and osmoprotective strategies to re-balance growth and physio-biochemical attributes in herbicidestressed Trigonella foenum-graecum. J Soil Sci Plant Nutr. 2021;21:3254-68.
- Sohag AAM, Tahjib-UI-Arif M, Brestic M, Afrin S, Sakil MA, Hossain MT, Hossain MA. Exogenous salicylic acid and hydrogen peroxide attenuate drought stress in rice. Plant Soil Environ. 2020;66(1):7-13.
- 32. Huang X, Tanveer M, Min Y, Shabala S. Melatonin as a regulator of plant ionic homeostasis: implications for abiotic stress tolerance. J Exp Bot. 2022;73(17):5886-902.
- Yan F, Zhang J, Li W, Ding Y, Zhong Q, Xu X, Wei H, Li G. Exogenous melatonin alleviates salt stress by improving leaf photosynthesis in rice seedlings. Plant Physiol Biochem. 2021;163:367-75.
- 34. Du Y, Zhao Q, Chen L, Yao X, Xie F. Effect of drought stress at reproductive stages on growth and nitrogen metabolism in soybean. Agronomy. 2020;10(2):302.
- Castañeda V, González EM. Strategies to apply water-deficit stress: similarities and disparities at the whole plant metabolism level in Medicago truncatula. Int J Mol Sci. 2021;22(6):2813.
- Heath RL, Packer L. Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. Arch Biochem Biophys. 1968;125(1): 189-98.
- 37. Kaya C, Shabala S. Melatonin improves drought stress tolerance of pepper (*Capsicum annuum*) plants via upregulating nitrogen metabolism. Funct Plant Biol. 2023;51(1).

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